

PROSTAGLANDIN-LIKE FATTY ACID DERIVATIVE FROM *CHROMOLAENA MORII*

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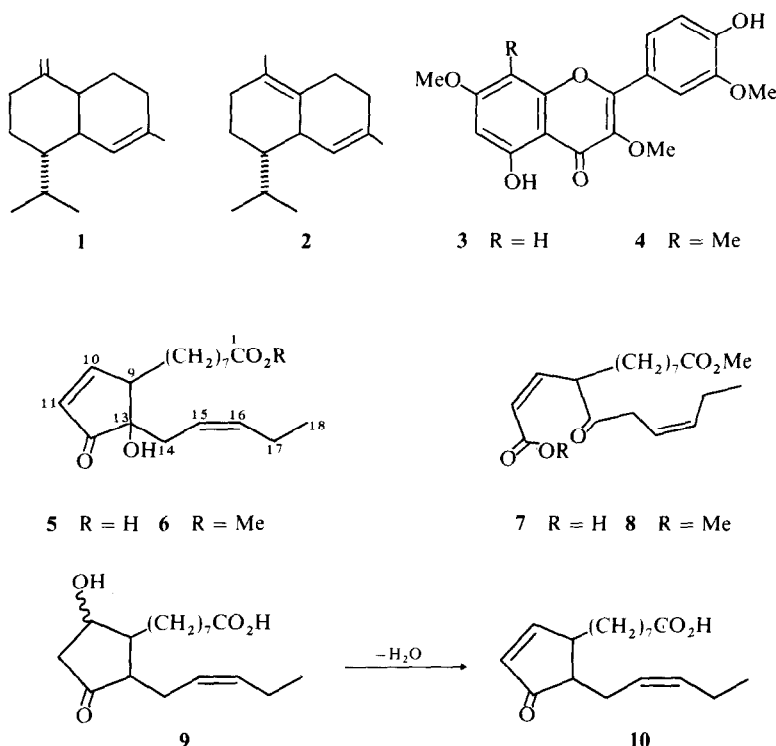
Key Word Index—*Chromolaena morii*; Compositae; cyclic fatty acid derivative.

The aerial parts of *Chromolaena morii* K. et. R. (tribe Eupatorieae, Compositae) afforded germacrene D, γ -humulene, caryophyllene, bicyclogermacrene, squalene, the cadinenes **1** and **2**, the flavonols **3** [1] and **4** [2] as well as the acid **5**, which was purified as its methyl ester **6**. Periodate cleavage of **6** gave the keto acid **7**, which was transformed to the dimethyl ester **8**. Most of the ^1H NMR data of **5** and its derivatives could be assigned by spin decoupling. The presence of the sequence H-14 to H-18 followed from the results of the decoupling of the signals of the olefinic protons ($\delta = 5.58$ and 5.32). Irradiation at 2.86 ppm decoupled the signals of H-10 and H-11, which showed a 6 Hz vicinal coupling with each other indicating a double bond in a five-membered ring. The position of the keto group followed from the downfield shift of the 10-H signal, while the position of the hydroxyl group could be deduced indirectly by the missing couplings of 14-H. These assignments were established by the ^1H NMR spectrum of **8**, which allowed a clear decision to be made as to the relative position of the substituents at C-9 and C-

13. The ^{13}C NMR data of **6** also supported the structure, while the mass spectrum showed the expected fragments: $\text{M} - \cdot\text{CH}_2\text{CH}=\text{CHCH}_2\text{Me}$ and $\text{M} - \cdot(\text{CH}_2)_7\text{CO}_2\text{Me}$. The configurations at C-9 and C-13 could not be determined. We have named **5** chromomoric acid. Probably **5** is formed from linolenic acid, in a similar way to the formation of the prostaglandins from arachidonic acid, through **9** and **10**. The latter has been reported as a product from the incubation of linolenic acid with a flax-seed extract [3]. The roots gave germacrene D, γ -humulene, caryophyllene and bicyclogermacrene. Only the cadinenes **1** and **2** may be typical for a *Chromolaena* species. Several oxygenated derivatives have been isolated from other species (see [4] and references cited therein).

EXPERIMENTAL

The air-dried plant material (voucher RMK 8067) was extracted with Et_2O -petrol (1:2). The resulting extracts were separated by CC (Si gel) and TLC (Si gel). The roots (100g)



afforded 20 mg germacrene D, 20 mg γ -humulene, 10 mg caryophyllene and 10 mg bicyclogermacrene, while the aerial parts (450 g) gave 100 g germacrene D, 50 mg γ -humulene, 50 mg caryophyllene, 20 mg bicyclogermacrene, 200 mg squalene, 10 mg **1**, 10 mg **2**, 50 mg **3**, 100 mg **4** and 20 mg **5** (Et₂O-petrol, 3:1). **5** was obtained as a colourless gum, which was esterified with CH₂N₂ to give **6** as a colourless gum, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 3550 (OH), 1740 (CO₂R), 1720 (C=O); MS m/z (rel. int.) 322.214 (M⁺, 4) (C₁₉H₃₀O₄), 304 (M - H₂O, 4), 291 (M - 'OMe, 7), 253.144 (M - 'CH₂CH=CHEt, 61) (C₁₄H₂₁O₄), 221 (253 - MeOH, 100), 165 (M - '(CH₂)₇ CO₂Me, 28), 147 (165 - H₂O, 15), 69 (C₅H₉⁺, 41).

$$[\alpha]_{24}^{25} = \frac{589}{+67} \frac{578}{+69} \frac{546}{+80} \frac{436 \text{ nm}}{+146} \quad (c = 0.8, \text{CHCl}_3).$$

¹³C NMR (CDCl₃): δ 208.5 (C-12) 174.2 (C-1), 164.2 (C-10), 136.3 (C-15), 130.1 (C-16), 121.5 (C-11), 80.7 (C-13), 52.0 (OMe), 51.4 (C-9), 34.1, 32.5, 29.5, 29.1, 28.2, 27.9, 25.0, 20.7 (CH₂), 14.0 (C-18). To 5 mg **6** in 1 ml MeOH 15 mg NaIO₄ were added. After 2 hr, **7** (2 mg) was isolated by TLC(Et₂O) as a colourless gum, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 3500–2600, 1735 (CO₂H), 1735 (CO₂R), 1690 (C=CCO). Addition of CH₂N₂ gave **8**, colourless gum, MS m/z (rel. int.): 352 (M⁺, 1), 321 (M - 'OMe, 10), 283 (M - C₅H₉, 18), 251 (283 - MeOH, 72), 69 (C₅H₉⁺, 100).

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Table 1. ¹H NMR data of compounds **6–8** (270 MHz, CDCl₃, TMS as internal standard)

| | 6 | 7 | 8 |
|-------------------|---|------------------------------|------------------------------|
| H-2 | 2.31 <i>t</i> | 2.32 <i>t</i> | 2.32 <i>t</i> |
| H-3 } H-8 } | 1.65 <i>m</i> | 1.65 <i>m</i> | 1.65 <i>m</i> |
| H-4 } H-7 } | 1.36 <i>m</i> | 1.33 <i>m</i> | 1.33 <i>m</i> |
| H-9 | 2.86 <i>m</i> | 3.30 <i>m</i> | 3.29 <i>m</i> |
| H-10 | 7.54 <i>dd</i> | 6.3 <i>m</i> | 6.15 <i>dd</i> |
| H-11 | 6.18 <i>dd</i> | 6.1 <i>d</i> (<i>br</i>) | 5.93 <i>dd</i> |
| H-14 } H-14' } | 2.50 <i>dd</i> (<i>br</i>) } 2.14 <i>dd</i> (<i>br</i>) } | 3.3 <i>m</i> } | 3.29 <i>m</i> } |
| H-15 } H-16 } | 5.58 <i>ddd</i> (<i>br</i>) } 5.32 <i>dt</i> (<i>br</i>) } | 5.55 <i>m</i> } | 5.55 <i>m</i> } |
| H-17 | 2.04 <i>dq</i> (<i>br</i>) | 2.05 <i>dq</i> (<i>br</i>) | 2.03 <i>dq</i> (<i>br</i>) |
| H-18 | 0.96 <i>t</i> | 0.97 <i>t</i> | 0.98 <i>t</i> |
| OMe | 3.67 <i>s</i> | 3.67 <i>s</i> | 3.67 <i>q</i> |

J (Hz): **6**: 2, 3 = 7.5; 9, 10 = 2; 9, 11 = 2; 10, 11 = 6.5; 14, 14' = 14.5; 14, 15 = 8; 14', 15 = 6; 15, 16 = 11; 16, 17 = 17, 18 = 7; **8**: 9, 10 = 10; 9, 11 = 2; 10, 11 = 11.

REFERENCES

1. Fraser, A. W. and Lewis, J. R. (1973) *Phytochemistry* **12**, 1787.
2. Fukui, K. and Matsumoto, T. (1965) *J. Chem. Soc. Jpn. Pure Chem. Sect.* **86**, 1079.
3. Zimmermann, D. C. and Feng, P. (1978) *Lipids* **13**, 313.
4. Bohlmann, F., Zdero, C., King, R. M. and Robinson, H. (1979) *Phytochemistry* **18**, 1177.